Product Manual

Cell Contraction Assay

Catalog Number

CBA-201

24 assays

FOR RESEARCH USE ONLY Not for use in diagnostic procedures



Introduction

Wound healing comprises of three processes: epithelialization, connective tissue deposition, and contraction. The contraction process is believed to be mediated by specialized fibroblasts called myofibroblasts. Three-dimensional collagen gels have been widely used in fibroblast contraction studies.

There are several different culture models to study the ability of fibroblasts to reorganize and contract collagen matrices in vitro. In the floating contraction model, a freshly polymerized collagen matrix containing cells is released from the culture dish and allowed to float in culture medium, and contraction occurs in the absence of external mechanical load and without appearance of stress fibers in the cells. In the attached model, a polymerized collagen matrix containing cells remains attached to the culture dish during contraction. Mechanical tension develops during contraction, and cellular stress fibers assemble. The two-step model combines an initial period of attached matrix contraction leading to mechanical loading, followed by release of the matrices, resulting in mechanical unloading and further contraction as mechanical stress dissipates.

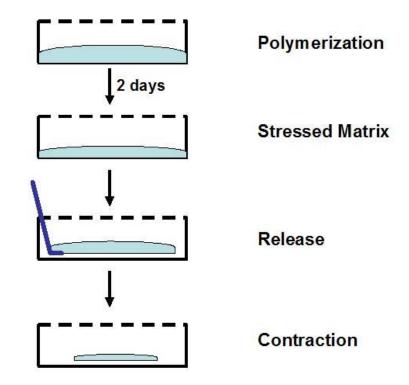
The signaling mechanisms used by fibroblasts to regulate collagen matrix contraction depend on whether the cells are mechanically loaded or unloaded at the time that contraction is initiated as well as on the growth factor used to initiate contraction. For instance, stimulation of fibroblasts by lysophosphatidic acid (LPA) but not by platelet-derived growth factor (PDGF) causes robust force generation in restrained matrices, whereas LPA and PDGF stimulate floating matrix contraction equally well.

3D collagen matrix has also been used in the studies of integrin signaling, cell apoptosis and cytoskeleton reorganization. Since three-dimensional matrix adhesions differ in structure, localization, and function from two-dimensional adhesions; and therefore, three-dimensional cell-matrix interactions may be more relevant biologically.

Cell Biolabs' Collagen-based Contraction Assay Kit provides a simple system to assess cell contractivity in vitro and screen cell contraction mediators. Each kit provides sufficient quantities to perform up to 24 assays in a 24-well plate. The kit can also be used for culturing cells in a 3D collagen matrix.



Assay Principle



Kit Components

- 1. <u>Collagen Solution</u> (Part No. 20101): One 10 mL bottle of sterile bovine Type I Collagen at 3.0 mg/mL
- 2. Neutralization Solution (Part No. 20102): One 0.5 mL tube
- 3. 5X DMEM Medium (Part No. 20103): One 5 mL bottle
- 4. <u>5X PBS (Part No. 20104)</u>: One 5 mL bottle
- 5. <u>100X Cell Contraction Inhibitor</u> (Part No. 20105): One 1 mL tube of 1M 2, 3-Butanedione Monoxime (BDM) in DMSO

Materials Not Supplied

- 1. Cells such as Fibroblast
- 2. Cell culture medium
- 3. 37°C Incubator, 5% CO2 atmosphere
- 4. Sterile Spatula
- 5. Light microscope
- 6. Ruler



Storage

Store all components at 4°C until their expiration dates.

Preparation of Reagents

Collagen Solution: The chart below is suggested for samples in 24-well plate, and may be modified accordingly to suit other culture plate sizes. Keep all solutions ON ICE all the time.

- 1. In a cold sterile tube, add desired amount of collagen solution. Next, add 5X DMEM medium or 5X PBS to the tube, mix well.
- 2. Add Neutralization solution, IMMEDIATELY mix and keep the collagen gel solution on ice.

Reagents	6 wells	12 wells	24 wells
Collagen Solution	2.385 mL	4.77 mL	9.54 mL
5X Medium or PBS	615 μL	1.23 mL	2.46 mL
Neutralization			
Solution	85 μL	170 μL	340 μL
Total	3.085 mL	6.17 mL	12.34 mL

Assay Protocol (Two-Step Collagen Contraction Model)

- 1. Harvest cells and resuspend in desired medium at $2-5 \times 10^6$ cells/mL.
- 2. Prepare collagen lattice by mixing 2 parts of cell suspension and 8 parts of cold collagen gel solution.
- 3. Add 0.5 mL of the cell-collagen mixture per well in a 24-well plate, incubate 1 hr at 37°C.
- 4. After collagen polymerization, 1.0 mL of culture medium is added atop each collagen gel lattice.
- 5. Cultures are incubated for two days, during which stress developed. Before release the stressed matrix, cells may be treated with contraction mediators, such as 10 mM BDM. To initiate contraction, gently release collagen gels from the sides of the culture dishes with a sterile spatula.
- 6. The collagen gel size change (contraction index) can be measured at various times with a ruler or quantified with image analysis software, such as NIH Image, Image Pro Plus.



Example of Results

The following figure demonstrates typical contraction results using the Cell Contraction Assay. One should use the data below for reference only. This data should not be used to interpret actual results.

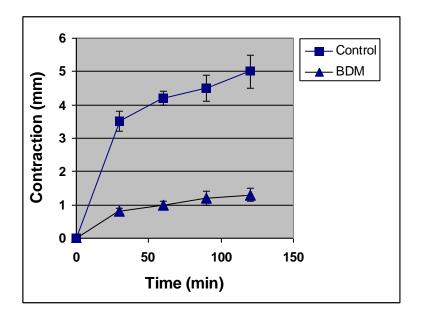


Figure 1. Contraction inhibition by BDM. 0.5×10^6 COS-7 cells in 0.5 mL collagen gel lattice were cultured for two days. Before initiation of contraction, cells were pretreated with 10 mM BDM for 1 hr. The change of gel size (diameter) in millimeters was measured with a ruler at various times after release.

References

- 1. Martin, P. (1997) Science 276, 75-81
- 2. Bell, E., Ivarsson, B., and Merrill, C. (1979) Proc. Natl. Acad. Sci. U. S. A. 76, 1274-1278
- 3. Stupak, D., and Harris, A. K. (1982) Dev. Biol. 90, 383-398
- 4. Mochitate, K., Pawelek, P., and Grinnell, F. (1991) Exp. Cell Res. 193, 198-207
- 5. Tian, B., Lessan, K., Kahm, J., Kleidon, J., and Henke, C. (2002) J. Biol. Chem. 277, 24667-24675

Recent Product Citations

- 1. Chen, P.Y. et al. (2016). Smooth muscle FGF/TGFβ cross talk regulates atherosclerosis progression. *EMBO Mol Med.* doi:10.15252/emmm.201506181.
- 2. Lazar-Karsten, P. et al. (2016). Generation and characterization of vascular smooth muscle cell lines derived from a patient with a bicuspid aortic valve. *Cells*. doi:10.3390/cells5020019.
- 3. Ye, Y. et al. (2016). Down-regulation of 14-3-3 Zeta inhibits TGF-β1–induced actomyosin contraction in human trabecular meshwork cells through rhoa signaling pathway14-3-3 zeta regulates actomyosin contraction in TM cells. *Invest Ophthalmol Vis Sci.* **57**:719-730.
- 4. Rinella, L. et al. (2016). Extracorporeal shockwaves modulate myofibroblast differentiation of adipose-derived stem cells. *Wound Repair Regen*. doi:10.1111/wrr.12410.



- 5. Halim, D. et al. (2015). ACTG2 variants impair actin polymerization in sporadic Megacystis Microcolon Intestinal Hypoperistalsis Syndrome. *Hum Mol Genet*. doi:10.1093/hmg/ddv497.
- 6. Li, H. Y. et al. (2015). Activation of TGF-β1-CD147 positive feedback loop in hepatic stellate cells promotes liver fibrosis. *Sci Rep.* **5**:16552.
- 7. Ham, S. A. et al. (2015). Ligand-activated PPARδ upregulates α-smooth muscle actin expression in human dermal fibroblasts: a potential role for PPARδ in wound healing. *J Dermatol Sci*. doi:10.1016/j.jdermsci.2015.10.005
- 8. Duru, N. et al. (2015). NRF2/miR-140 signaling confers radioprotection to human lung fibroblasts. *Cancer Lett.* doi:10.1016/j.canlet.2015.08.011.
- 9. Gutiérrez, J. et al. (2015). RECK-mediated β1-integrin regulation by TGF-β1 is critical for wound contraction in mice. *PLoS One.* **10**:e0135005.
- 10. Ratuszny, D. et al. (2015). miR-145 is a promising therapeutic target to prevent cornea scarring. *Hum Gene Ther*. doi:10.1089/hum.2014.151.
- 11. Sobel, K. et al. (2015). FTY720-P activates sphingosine-1-phosphate receptor 2 and selectively couples to Gα12/13/Rho/ROCK to induce myofibroblast contraction. *MolPharmacol*. doi:10.1124/mol.114.097261.
- 12. Woeller, C. F. et al. (2015). Salinomycin and other polyether ionophores are a new class of antiscarring agent. *J Biol Chem.* **290**:3563-3575.
- 13. Luo, Y. et al. (2015). Magnitude-dependent proliferation and contractility modulation of human bladder smooth muscle cells under physiological stretch. *World J Urol.* doi:10.1007/s00345-015-1509-4.
- 14. Reddy, A. T. et al. (2014). Nitrated fatty acids reverse pulmonary fibrosis by dedifferentiating myofibroblasts and promoting collagen uptake by alveolar macrophages. *FASEB J.* **28**:5299-5310.
- 15. Rubattu, S. et al. (2014). The C2238/αANP variant is a negative modulator of both viability and function of coronary artery smooth muscle cells. *PLoS One.* **17**:e113108.
- 16. Hsieh, Y. P. et al. (2014). Arecoline stimulated early growth response-1 production in human buccal fibroblasts: Suppression by epigallocatechin-3-gallate. *Head Neck*. doi:10.1002/hed.23614.
- 17. Je, Y. J. et al. (2014). Inhibitory role of Id1 on TGF-β-induced collagen expression in human dermal fibroblasts. *Biochem Biophys Res Commun.* **444**:81-85.
- 18. Herr, M. J. et al. (2014). Tetraspanin CD9 regulates cell contraction and actin arrangement via rhoa in human vascular smooth muscle cells. *PLoS One*. **9**:e106999.
- 19. Nie, L. et al. (2014). Endothelial-mesenchymal transition in normal human esophageal endothelial cells cocultured with esophageal adenocarcinoma cells: role of IL-1β and TGF-β2. *Am J Physiol Cell Physiol.* **307**:C859-C877.
- 20. Kotio, K.U. et al. (2011). Implication of microRNAs in Atrial Natriuretic Peptide and Nitric Oxide Signaling in Vascular Smooth Muscle Cells. *Am J Physiol Cell Physiol.* **301**:C929-C937.
- 21. Schell, C. et al. (2010). 15-deoxy-delta12-14-prostaglandin-J2 induces hypertrophy and loss of contractility in human testicular peritubular cells: implications for human male fertility. *Endocrinology* **151**:1257-1268.

Warrantv

These products are warranted to perform as described in their labeling and in Cell Biolabs literature when used in accordance with their instructions. THERE ARE NO WARRANTIES THAT EXTEND BEYOND THIS EXPRESSED WARRANTY AND CELL BIOLABS DISCLAIMS ANY IMPLIED WARRANTY OF MERCHANTABILITY OR WARRANTY OF FITNESS FOR PARTICULAR PURPOSE. CELL BIOLABS' sole obligation and purchaser's exclusive remedy for breach of this warranty shall be, at the option of CELL BIOLABS, to repair or replace the products.



In no event shall CELL BIOLABS be liable for any proximate, incidental or consequential damages in connection with the products.

Contact Information

Cell Biolabs, Inc. 7758 Arjons Drive San Diego, CA 92126 Worldwide: +1 858-271-6500

USA Toll-Free: 1-888-CBL-0505 E-mail: tech@cellbiolabs.com

www.cellbiolabs.com

©2004-2016: Cell Biolabs, Inc. - All rights reserved. No part of these works may be reproduced in any form without permissions in writing.

